





Draft Genome Sequence of Bryobacteraceae Strain F-183

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ABSTRACT Here, we report a draft genome sequence of a bacterial strain, F-183, isolated from a duckweed frond. Strain F-183 belongs to the family *Bryobacteraceae* of the phylum *Acidobacteria*, and its genomic information would contribute to understanding the ecophysiology of this abundant but rarely characterized phylum.

The phylum *Acidobacteria* is one of the most abundant and widespread bacterial groups in soils and is indeed distributed widely across environments such as terrestrial plants, hot springs, mine water, sediments, and marine sponges (1, 2). However, only 61 species have been validly described in this phylum at present despite its wide distribution and phylogenetic diversity based on the 16S rRNA gene phylogeny (1, 2), hampering characterization of its physiology and ecological roles in various natural environments.

Strain F-183 had been isolated previously from fronds of wild duckweeds collected from a pond located in Tsukuba City, Ibaraki, Japan (3). DNA extraction was performed with a procedure reported previously (4). Briefly, genomic DNA was extracted from cells cultivated with PE03 medium (3) at 30°C under shaking conditions by digestion using lysozyme, sodium dodecyl sulfate, and proteinase K, followed by purification by phenol-chloroform extraction and ethanol precipitation. For extraction of high-molecular-weight (HMW) DNA, the MagAttract HMW DNA kit (Qiagen) was used according to the manufacturer's instruction.

Library preparation and sequencing were performed by using commercial kits according to the manufacturer's instructions (Table 1). A total of 2.56 million of paired-end reads and 3.28 million of mate-pair reads (Illumina MiSeq) and 0.39 million of single long reads (mean length, 8,914 bp) with the MinION system (Oxford Nanopore Technologies) were obtained. Read quality control was performed by FastQC version 0.11.5 (5). The obtained sequence data were assembled using hybridSPAdes version 3.13.0 (6) in KBase (7). Genome annotation was performed using the DDBJ Fast Annotation and Submission Tool (DFAST) pipeline version 1.2.13 (8). Structural annotations were performed using MetaGeneAnnotator version 2008/08/19 (9) for CDS, Barrnap version 0.8 (10) for rRNA, ARAGORN version 1.2.38 (11) for

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TABLE 1 Library preparation and sequencing

	Equipment by sequencing type	
Procedure		MinION long-read sequencing
Library preparation	KAPA HyperPrep kit (for Illumina) (Kapa Biosciences); Illumina Nextera mate-pair	Rapid sequencing kit (Oxford
	library prep kit (Illumina); insert length for mate-pair libraries were 3 kb and 8 kb	Nanopore Technologies)
Sequencing platform	Illumina MiSeq system (paired end, 2 $ imes$ 300 bp)	MinION system (R9.4 flow cell)

tRNA, and CRT version 1.2 (12) for CRISPR. Genome completeness was estimated with CheckM version 1.1.3 (13). Taxonomic assignment was performed using the Genome Taxonomy Database Toolkit (GTDB-Tk) version 0.1.4 (14). Default parameters were used for all software.

One scaffold sequence having a single assembly gap and three short contigs were generated by the hybrid assembly. The circular structure of each sequence was verified by Sanger sequencing, and the overlap sequences were trimmed. Finally, the F-183 nearly complete genome was recovered at $99\times$ coverage and consists of one circular chromosome (N_{50} , 6,182,012 bp) and three circular sequences (45,950, 40,882, and 12,955 bp) with a total G+C content of 60.1%. The genome contains 5,539 protein-coding sequences, 49 tRNA genes, and 3 rRNAs. No CRISPRs were detected. The genome was determined to be 95.46% complete and 3.65% redundant and to have 0% strain heterogeneity. Strain F-183 was placed within the family *Bryobacteraceae* of the phylum *Acidobacteriota* (*Acidobacteria*) but was not assigned to a genus.

As strain F-183 exhibits the ability to promote plant growth (3), a further genome analysis would be helpful for understanding the mechanisms of F-183 interactions with aquatic plants and its physiology and ecological function.

Data availability. The genome and raw sequences have been deposited in DDBJ/ENA/GenBank under accession numbers AP025252, AP025253, AP025254, and AP025255 and in the DDBJ Sequence Read Archive under accession numbers DRA011790 and DRA013042.

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REFERENCES

- Kielak AM, Barreto CC, Kowalchuk GA, van Veen JA, Kuramae EE. 2016. The ecology of Acidobacteria: moving beyond genes and genomes. Front Microbiol 7:744. https://doi.org/10.3389/fmicb.2016.00744.
- Kalam S, Basu A, Ahmad I, Sayyed RZ, El-Enshasy HA, Dailin DJ, Suriani NL. 2020. Recent understanding of soil acidobacteria and their ecological significance: a critical review. Front Microbiol 11:580024. https://doi.org/10 .3389/fmicb.2020.580024.
- Yoneda Y, Yamamoto K, Makino A, Tanaka Y, Meng X-Y, Hashimoto J, Shin-Ya K, Satoh N, Fujie M, Toyama T, Mori K, Ike M, Morikawa M, Kamagata Y, Tamaki H. 2021. Novel plant-associated acidobacteria promotes growth of common floating aquatic plants, duckweeds. Microoganisms 9:1133. https://doi.org/10.3390/microorganisms9061133.
- Komatsu M, Komatsu K, Koiwai H, Yamada Y, Kozone I, Izumikawa M, Hashimoto J, Takagi M, Omura S, Shin-Ya K, Cane DE, Ikeda H. 2013. Engineered Streptomyces avermitilis host for heterologous expression of biosynthetic gene cluster for secondary metabolites. ACS Synth Biol 2: 384–396. https://doi.org/10.1021/sb3001003.
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. http://www.bioinformatics.babraham.ac.uk/projects/fastqc/.
- 6. Antipov D, Korobeynikov A, McLean J, Pevzner P. 2016. hybridSPAdes: an

- algorithm for hybrid assembly of short and long reads. Bioinformatics 32: 1009–1015. https://doi.org/10.1093/bioinformatics/btv688.
- 7. Arkin AP, Cottingham RW, Henry CS, Harris NL, Stevens RL, Maslov S, Dehal P, Ware D, Perez F, Canon S, Sneddon MW, Henderson ML, Riehl WJ, Murphy-Olson D, Chan SY, Kamimura RT, Kumari S, Drake MM, Brettin TS, Glass EM, Chivian D, Gunter D, Weston DJ, Allen BH, Baumohl J, Best AA, Bowen B, Brenner SE, Bun CC, Chandonia J-M, Chia J-M, Colasanti R, Conrad N, Davis JJ, Davison BH, DeJongh M, Devoid S, Dietrich E, Dubchak I, Edirisinghe JN, Fang G, Faria JP, Frybarger PM, Gerlach W, Gerstein M, Greiner A, Gurtowski J, Haun HL, He F, Jain R, et al. 2018. KBase: the United States Department of Energy Systems Biology Knowledgebase. Nat Biotechnol 36:566–569. https://doi.org/10.1038/nbt.4163.
- 8. Tanizawa Y, Fujisawa T, Nakamura Y. 2018. DFAST: a flexible prokaryotic genome annotation pipeline for faster genome publication. Bioinformatics 34:1037–1039. https://doi.org/10.1093/bioinformatics/ btx713.
- Noguchi H, Taniguchi T, Itoh T. 2008. MetaGeneAnnotator: detecting species-specific patterns of ribosomal binding site for precise gene prediction in anonymous prokaryotic and phage genomes. DNA Res 15:387–396. https://doi.org/10.1093/dnares/dsn027.

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- 10. Seemann T. 2013. Barrnap 0.7: rapid ribosomal RNA prediction.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res 32:11–16. https://doi.org/10.1093/nar/gkh152.
- Bland C, Ramsey TL, Sabree F, Lowe M, Brown K, Kyrpides NC, Hugenholtz P. 2007. CRISPR Recognition Tool (CRT): a tool for automatic detection of clustered regularly interspaced palindromic repeats. BMC Bioinformatics 8:209. https://doi.org/10.1186/1471-2105-8-209.
- 13. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. Genome Res 25:1043–1055. https://doi.org/10.1101/gr.186072.114.
- 14. Chaumeil P-A, Mussig AJ, Hugenholtz P, Parks DH. 2019. GTDB-Tk: a toolkit to classify genomes with the Genome Taxonomy Database. Bioinformatics 36:1925–1927. https://doi.org/10.1093/bioinformatics/btz848.